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Title of Paper: Paeoniflorin Protects Against Retinal Photoreceptor Degeneration in a Mouse Model of Light-induced Retinal Injury

IRB Status: Approved N/A

Type of Study: Basic science Case report, series Retrospective Prospective
 Randomized Controlled

If the project was previously presented at a major ophthalmology meeting (e.g. ARVO, AAO, ASCRS), please indicated date of presentation(s):
ARVO - May 2017

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1 **Paeoniflorin Protects Against Retinal Photoreceptor Degeneration in a Mouse Model of Light-induced**
2 **Retinal Injury**

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31 Abstract**32 Purpose:**

33 Age-related macular degeneration (AMD) is a leading cause of blindness in the United States and other
34 developed nations. Current treatment modalities targeting AMD are beneficial to only a small proportion
35 of patients with the end-stage form and do not necessarily prevent progression of the disease.

36 Paeoniflorin (PF) is one of the main active components of *Paeonia Radix*, a traditional Chinese herbal
37 medicine derived from the root of *Paeonia lactiflora* Pallas. Paeoniflorin possesses both anti-inflammatory
38 and anti-oxidant properties and has been implicated as a potential therapy in other degenerative
39 pathologies such as Alzheimer's disease. We hypothesize that paeoniflorin will have protective effects
40 against retinal degeneration in a mouse model of retinal light injury.

41 Methods:

42 BALB/c albino mice were assigned to one of two groups: control-treated or paeoniflorin-treated retinal
43 light injury. Control (PBS) or paeoniflorin (20 mg/kg in PBS) was injected intraperitoneally and mice were
44 then exposed to 10,000 lux cool white fluorescent light for 2 hours to induce light injury. Both groups had
45 treatment administered once daily for 4 additional days following light injury for a total of 5 treatments.
46 Scotopic electroretinography (ERG) was recorded before light injury and 7 days following light injury.
47 After ERG recording 7 days following light injury, mice eyes were then enucleated and fixed in phosphate-
48 buffered (pH 7.4) solution of 2.5% glutaraldehyde-2% paraformaldehyde. The eye cups were dissected
49 and then embedded in epoxy resin for retinal histology and morphometry.

50 Results:

51 Before treatment, scotopic ERG a- and b-wave amplitudes were an average of $542 \pm 31 \mu\text{V}$ and 1005 ± 61
52 μV , respectively, in the control group and $452 \pm 109 \mu\text{V}$ and $867 \pm 147 \mu\text{V}$, respectively, in the
53 paeoniflorin treatment group. Following retinal light injury, ERG a-wave amplitudes were $132 \pm 17 \mu\text{V}$ in
54 the control group and $250 \pm 59 \mu\text{V}$ in the paeoniflorin treatment group ($p < 0.005$). Following retinal light
55 injury, ERG b-wave amplitudes were $304 \pm 42 \mu\text{V}$ in the control group and $518 \pm 138 \mu\text{V}$ in the
56 paeoniflorin treatment group ($p < 0.005$). Baseline a- and b-wave amplitudes between control and
57 treated groups were statistically indistinguishable ($p > 0.05$). In control treated mice, light injury produced
58 significant thinning of the outer nuclear layer (ONL), but the degree of thinning of the ONL was
59 significantly less in cases of paeoniflorin treatment. There was significantly greater preservation of ERG a-
60 and b-wave amplitudes in the paeoniflorin treatment group as well as greater histologic preservation of
61 retinal layers.

62 Conclusion:

63 Paeoniflorin treatment of mice had a significant impact on the preservation of ERG a-and b-wave
64 amplitudes following retinal light injury when compared to a control group. Greater histologic
65 preservation was also seen with paeoniflorin treatment. Our preliminary findings suggest that
66 paeoniflorin may have therapeutic value in the management of retinal degenerative conditions.

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86 Introduction

87 AMD is a leading cause of blindness in the developed world. In the United States alone, an estimated 10
88 million Americans are affected by AMD. Current treatment modalities targeting AMD are beneficial to
89 only a small proportion of patients with end-stage disease. Furthermore, FDA-approved treatments only
90 exist for exudative “wet, neovascular” AMD, not for the non-exudative “dry, atrophic” form.¹ An
91 incomplete understanding of its pathogenesis has limited progress in the development of novel
92 therapeutics against this disease. Although mainstay treatment options include high-dose antioxidant
93 vitamins, both the AREDS and AREDS2 formulations mainly have been shown to reduce the incidence of
94 neovascular AMD in select individuals. No convincing data exist to suggest either of these antioxidant
95 formulations prevent the progression to chronic geographic atrophy, the severe end-stage form of dry
96 AMD.²

97 Paeoniflorin is one of the main active components of *Paeonia Radix*, a traditional Chinese herbal medicine
98 derived from the root of *Paeonia lactiflora* Pall. Previous studies have demonstrated a protective role of
99 paeoniflorin in a cell culture model of AMD.³ Paeoniflorin possesses both anti-inflammatory and anti-
100 oxidant properties and has also been implicated as a potential therapy in other degenerative pathologies
101 such as Alzheimer’s disease.^{3,4,5} However, no studies to date have examined the therapeutic potential of
102 paeoniflorin in a live animal model of retinal degeneration. Furthermore, the potential protective effect
103 of this compound specifically on retinal photoreceptors, a key cellular structure that is affected in AMD,
104 has not been investigated.

105 In this study we employ a live mouse model of retinal light injury to investigate the potential protective
106 benefits of paeoniflorin against this specific type of retinal degeneration. The mouse retinal light-induced
107 injury model has been successfully used in retinal neurodegenerative disease studies.⁶ Oxidative stress, in
108 particular, has been shown to be an important mechanism involved in light-induced retinal damage.⁷
109 Although mice anatomically lack a macula, other components such as the neuroretina and the retinal
110 pigment epithelium can develop lesions that portray clinical features of AMD and have been widely used
111 in studies of potential therapeutics against this vision-threatening condition. Shibagaki, K et al. recently
112 employed a mouse retinal light-induced injury model to investigate the potential benefits of another
113 compound against oxidative stress in such diseases such as AMD. In that study, the compound of interest,
114 pramipexole, was found to have a protective benefit in the animal model.⁸ Studies have also confirmed
115 that exposure of the retina to increased levels of oxidative stress in animal models leads to loss of
116 photoreceptors and retinal pigment epithelium, two main hallmarks of AMD.⁹ Given the previously

117 described anti-inflammatory and anti-oxidant properties of paeoniflorin, we suggest paeoniflorin may
118 have therapeutic value in the management of retinal degenerative conditions.

119

120 **Materials and Methods**

121 *IACUC Statement*

122 All procedures involving mice were performed according to the ARVO statement for the use of Animal in
123 Ophthalmic and Vision Research and were approved by the Institutional Animal Care and Use Committee
124 of Edward Hines, Jr. VA Hospital.

125 *Light-induced retinal injury*

126 All exposure to light began at 10 AM. The BALB/c albino mice (8 weeks) were dark adapted for 24 hours
127 before the experiments, and the pupils were dilated with 1% cyclopentolate hydrochloride eye drops 1
128 hour before exposure to light and right before exposure to the light. Non-anesthetized mice were
129 exposed to 10,000 lux of diffuse, cool, white fluorescent light for 2 hours (from 10AM-12AM). A special
130 device was used with round light bulbs to prevent the mice hiding their face. The temperature during
131 exposure to light was maintained at $25 \pm 1.5^{\circ}\text{C}$.

132 *Paeoniflorin Treatment*

133 Paeoniflorin was dissolved in PBS (stock 200 mg/ml). Paeoniflorin (20 mg/kg/day) or control (PBS) was
134 administered one time before light injury and for 4 additional days following retinal light injury via
135 intraperitoneal injection.

136 *Electroretinography (ERG)*

137 Retinal function (ERG) were measured before and 7 days after inducing retinal light injury. Mice were
138 dark-adapted overnight, anesthetized with ketamine (100mg/kg) and xylazine (5mg/kg) and their pupils
139 dilated with 1% tropicamide and 2.5% phenylephrine hydrochloride. Using a stainless-steel electrode
140 coated with 1% methylcellulose, the ERG was recorded from the corneal surface with a series of stimulus
141 luminance. Needle electrodes were subcutaneously inserted in the cheek and the tail served as reference
142 and ground leads, respectively. The ERG responses were differentially amplified (0.3–1,500 Hz), averaged
143 and stored using a UTAS E-3000 signal averaging system (LKC Technologies, Gaithersburg, MD). A notch
144 filter at 60 Hz was used during recording. Stimuli ranged from -3.6 to $2.1 \log \text{cd s/m}^2$ and were presented

145 in increasing order in dark and at least two successive responses were averaged together for each
146 stimulus presented. Intervals between the stimuli were increased from 4 s to 61 s. The body temperature
147 of anesthetized mice was kept at 37° C using a temperature-regulated heating pad.

148 *Histology and Morphometry*

149 The eyes from all groups were collected after ERG recording at 7 days after inducing retinal light injury.
150 Enucleated eyes were fixed in phosphate-buffered (pH 7.4) solution of 2.5% glutaraldehyde-2%
151 paraformaldehyde. The eye cups were dissected and then embedded in epoxy resin. Sections with 1 µm
152 thickness were cut vertically through the optic nerve and subsequently stained with toluidine blue. Three
153 separate morphometric measurements of the outer nerve layer (ONL) from the superior and inferior
154 retina between 750-1000 µm from the optic disc were averaged together. The histology changes were
155 evaluated by measuring the outer retinal thickness in a protocol previously described.¹⁰

156 *Statistical Analysis*

157 Statistical significance between the means of multiple experimental groups will be determined using one-
158 way analysis of variance (ANOVA). An F ratio giving a value of $p < 0.05$ is considered significant.

159 **Results**

160 Before treatment, scotopic ERG a- and b-wave amplitudes were an average of $542 \pm 31\mu\text{V}$ and 1005 ± 61
161 μV , respectively, in the control group and $452 \pm 109 \mu\text{V}$ and $867 \pm 147 \mu\text{V}$, respectively, in the
162 paeoniflorin treatment group (Figure 1 and 2). Following retinal light injury, ERG a-wave amplitudes were
163 $132 \pm 17 \mu\text{V}$ in the control group and $250 \pm 59 \mu\text{V}$ in the paeoniflorin treatment group ($p < 0.005$).
164 Following retinal light injury, ERG b-wave amplitudes were $304 \pm 42 \mu\text{V}$ in the control group and $518 \pm$
165 $138 \mu\text{V}$ in the paeoniflorin treatment group ($p < 0.005$). There was significantly greater preservation of
166 ERG a- and b-wave amplitudes in the paeoniflorin treatment group suggesting that paeoniflorin is able to
167 attenuate the degree of degenerative loss of retinal function in a retinal light injury model.

168 Light injury induced photoreceptor degeneration and photoreceptor cell death was evidenced in toluidine
169 blue stained sections after 7 days post retinal light injury. However, the thinning of the ONL in the
170 paeoniflorin-treated light injury group was relatively preserved as compared to that seen in the control-
171 treated light injury group (Figure 3). The difference in mean thickness of the ONL at 750 - 1000 µm from
172 the optic disc between the control and the paeoniflorin-treated light injury groups was statistically
173 significant (Figure 4).

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175 Discussion

176 Our findings demonstrate that paeoniflorin treatment can protect against retinal degeneration in a
177 mouse model of retinal light injury. This preventive effect was observed in both functional and structural
178 studies. The exact mechanism by which paeoniflorin exerts its potential protective effect is unclear.
179 However, as previously described, paeoniflorin possess both anti-inflammatory and anti-oxidative
180 properties.^{3,4,5} Given that prior studies have also confirmed that exposure of the retina to increased
181 levels of oxidative stress in animal models leads to loss of photoreceptors and retinal pigment epithelium,
182 which are two main hallmarks of AMD, it is not surprising that a compound with these properties is able
183 to produce findings suggestive of a protective effect against this type of degeneration. While paeoniflorin
184 has been demonstrated to have protective effects in other neurodegenerative conditions^{3,4,5}, we report
185 for the first time a similar effect of paeoniflorin on retinal function. Further research to elucidate the
186 exact mechanisms involved are warranted.

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188 Acknowledgments

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190 Prevention of Blindness.

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192 References

- 193 1. Ambati J, Fowler BJ. Mechanisms of age-related macular degeneration. *Neuron*. 2012;75:26–39.
- 194 2. The Age-Related Eye Disease Study 2 (AREDS2) Research Group, Chew EY, Clemons TE, et al. Secondary
195 Analyses of the Effects of Lutein/Zeaxanthin on Age-Related Macular Degeneration Progression AREDS2
196 Report No.3. *JAMA ophthalmology*. 2014;132(2):142-149. doi:10.1001/jamaophthalmol.2013.7376.
- 197 3. Wankun X, Wenzhen Y, Min Z, et al. Protective effect of paeoniflorin against oxidative stress in human
198 retinal pigment epithelium in vitro. *Molecular Vision*. 2011;17:3512-3522.
- 199 4. Wang QS, Gao T, Cui YL, Gao LN, Jiang HL. Comparative studies of paeoniflorin and albiflorin from paeonia
200 lactiflora on anti-inflammatory activities. *Pharm Biol*. 2014; 52:1189-1195. doi:
201 10.3109/13880209.2014.880490.

- 202 5. Liu H., Wang J., Wang J., Wang P., Xue Y. Paeoniflorin attenuates A β 1–42-induced inflammation and
203 chemotaxis of microglia *in vitro* and inhibits NF- κ B- and VEGF/Flt-1 signaling pathways. *Brain Res.*
204 2015;1618:149–158. doi: 10.1016/j.brainres.2015.05.035.
- 205 6. Oishi A, Otani A, Sasahara M, Kojima H, Nakamura H, et al. 2008. Granulocyte colony-stimulating factor
206 protects retinal photoreceptor cells against light-induced damage. *Invest Ophthalmol Vis Sci* 49:5629-35.
- 207 7. Wang S. et al. 17 β -Estradiol Ameliorates Light-Induced Retinal Damage in Sprague–Dawley Rats by
208 Reducing Oxidative Stress. *Journal of Molecular Neuroscience* 55, 141–151 (2015).
- 209 8. Shibagaki K., Okamoto K., Katsuta O., Nakamura M. Beneficial protective effect of pramipexole on light-
210 induced retinal damage in mice. *Experimental Eye Research*. 2015;139:64–72.
211 doi:10.1016/j.exer.2015.07.007.
- 212 9. Collier RJ, Wang Y, Smith SS, Martin E, Ornberg R, Rhoades K *et al.* Complement deposition and microglial
213 activation in the outer retina in light-induced retinopathy: inhibition by a 5-HT1A agonist. *Invest*
214 *Ophthalmol Vis Sci* 2011; 52(11): 8108–8116.
- 215 10. Bu P, Basith B, Stubbs EB, Jr., Perlman JI. 2010. Granulocyte colony-stimulating factor facilitates recovery of
216 retinal function following retinal ischemic injury. *Exp EyeRes* 91:104-6

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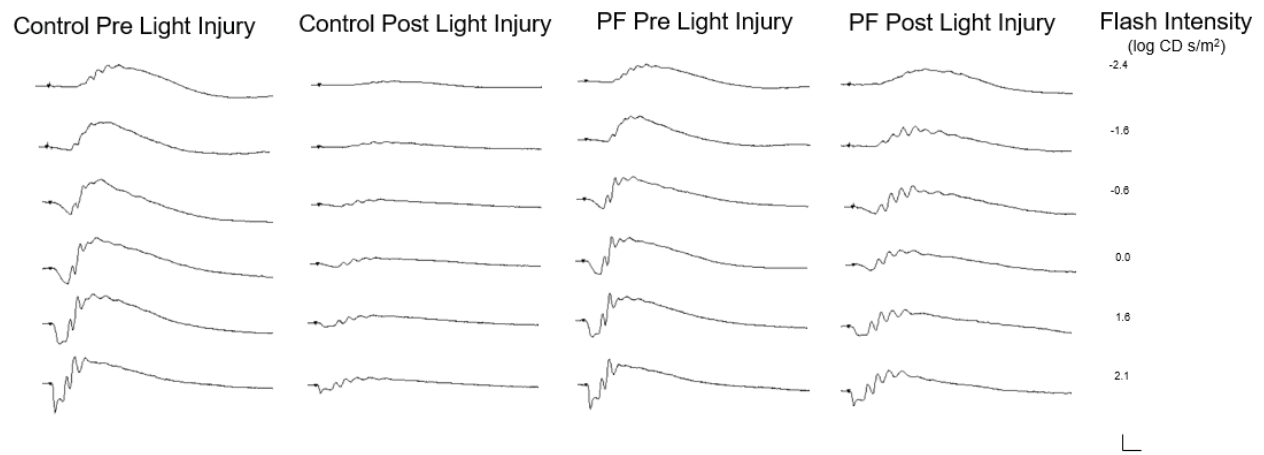
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232 Figures



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234 **Figure 1.** Representative dark-adapted ERG a- and b-waves. Preservation of a- and b-waves in the
 235 paeoniflorin treatment group compared to that of the control group is demonstrated. Scale bars: 20 ms
 236 (x-axis) and 250 μV (y-axis).

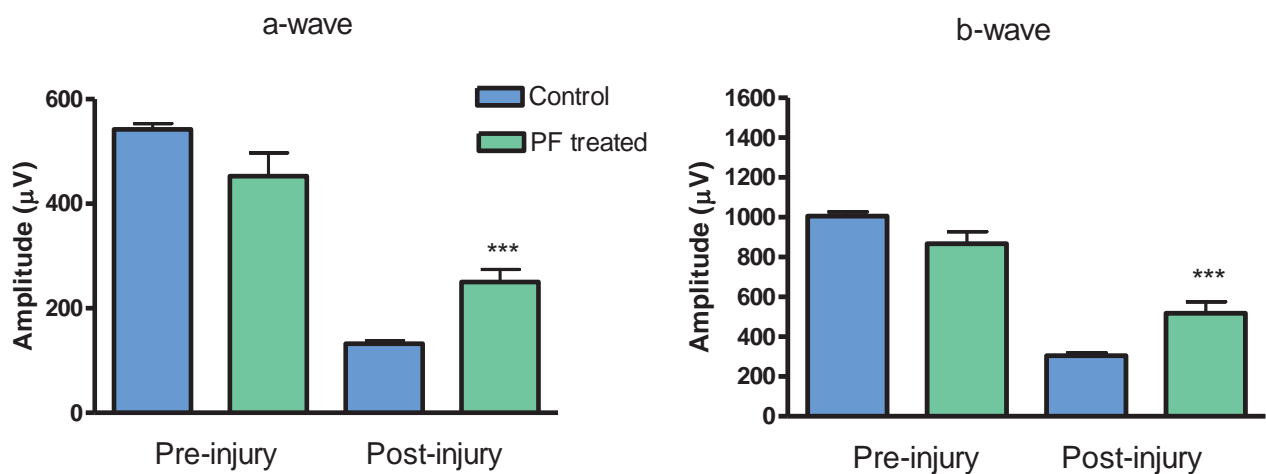
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243 **Figure 2.** Effect of paeoniflorin treatment on retinal function following light injury. Quantitative changes in
 244 ERG a- and b-wave amplitudes at 2.1 log CD s/m² flash intensity.

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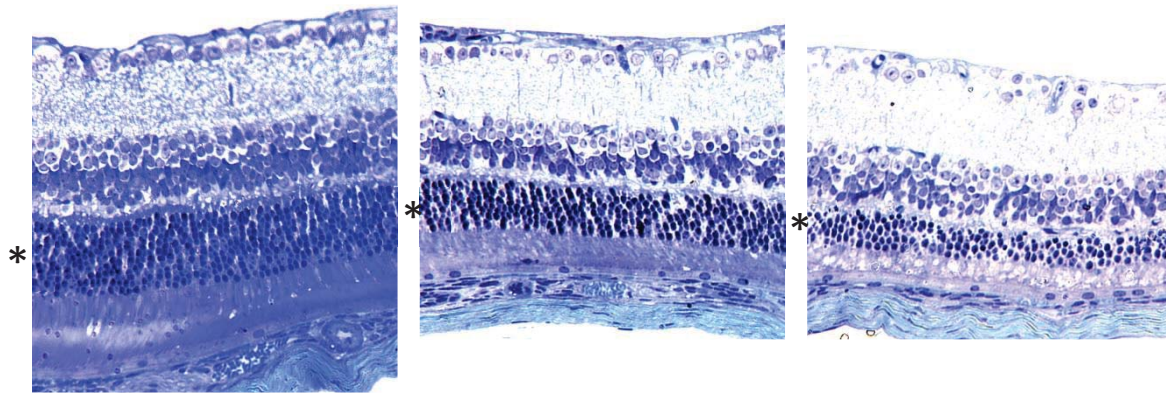
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No Light
Injury

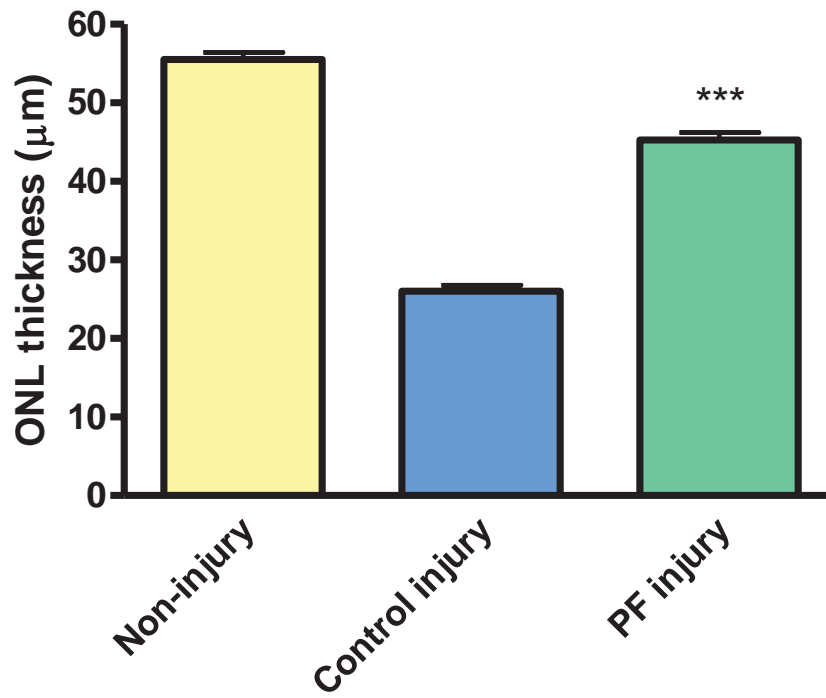
PF Light Injury

Control Light
Injury

256 **Figure 3.** Effect of paeoniflorin on retinal histology following retinal light injury. Representative stained
257 retinal sections from normal (No Light Injury), paeoniflorin-treated (PF Light Injury) or control-treated
258 (Control Light Injury) tissues. Data shown are representative samples (n = 3). An * is used to indicate the
259 outer nuclear layer.

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263 **Figure 4.** Morphometry data quantifying the preservation of the outer nuclear layer thickness between
264 the no light injury (non-injury), control-treated light injury (control injury) and the paeoniflorin-treated
265 light injury (PF injury) groups.

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